

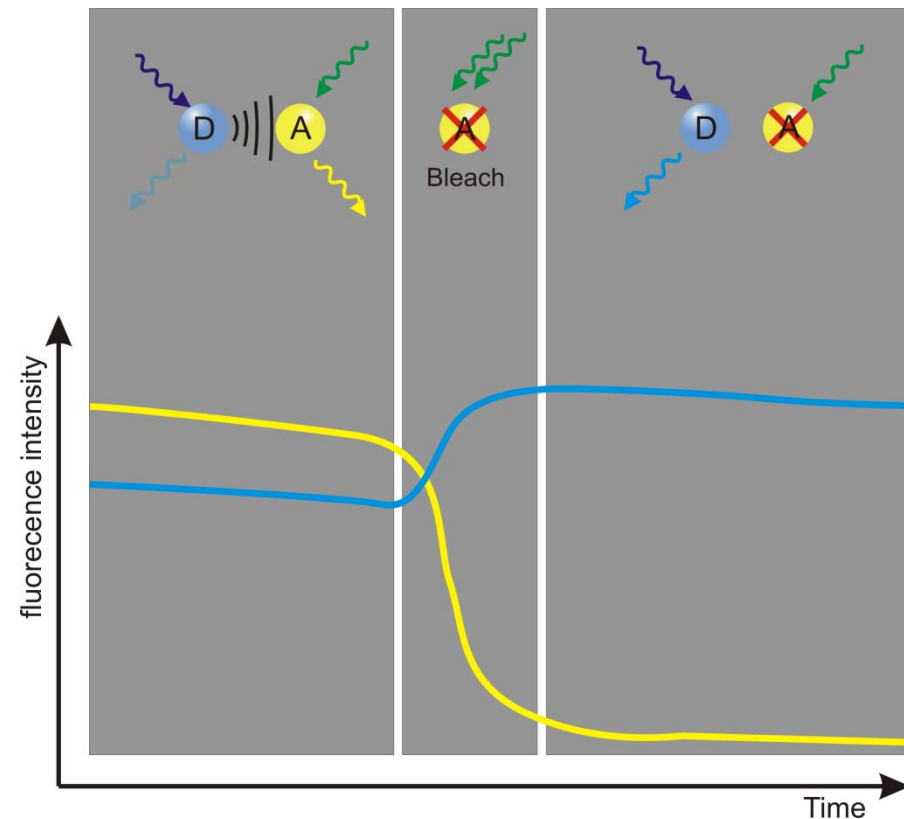
FRET Detection: Acceptor Photobleaching

Principle of Acceptor Photobleaching



Acceptor photobleaching: a simple FRET detection method

De-quenching of the donor after selective photobleaching of the acceptor causes an increase in donor emission that can be readily quantified.



FRET Detection: Acceptor Photobleaching

ZEN 2010: Image acquisition



(1) Imaging Setup

- Multitrack Channel Mode
Linewise switch (fast)
- 1) Excite Donor and Detect Donor emission
- 2) Excite Acceptor and Detect Acceptor emission



(2) Regions

- Select the cells / structures to be analyzed by defining bleach ROIs (ROIs = Regions of Interest)



(3) Bleaching

- Acquire a couple of pre-bleach images to enable the system to calculate average intensities for analysis (baseline)
- Instead of one-shot bleach define repetitive bleach events (maximum delta donor intensity)
- Start with a pre-experiment to test bleaching conditions: „Stop when intensity drops to XX %“
- Apply high laser power / iteration
- Bleach acceptor to ~ 10-20% of its initial value



(4) Time Series

- Acquire a number of time points

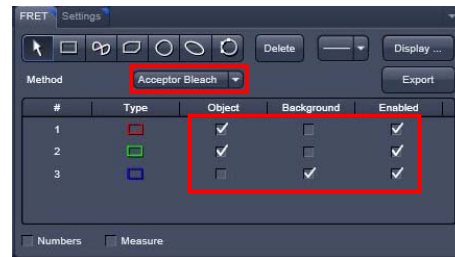
FRET Detection: Acceptor Photobleaching

ZEN 2010: Data evaluation using FRET



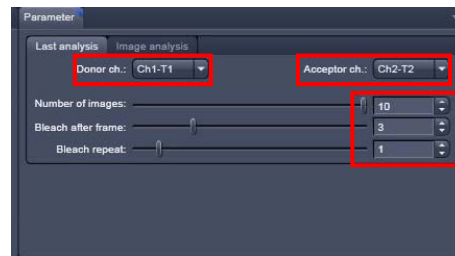
(1) FRET

- Method: Acceptor Bleach
- Define ROIs: Enable Object(s) and Background
- NO reference possible



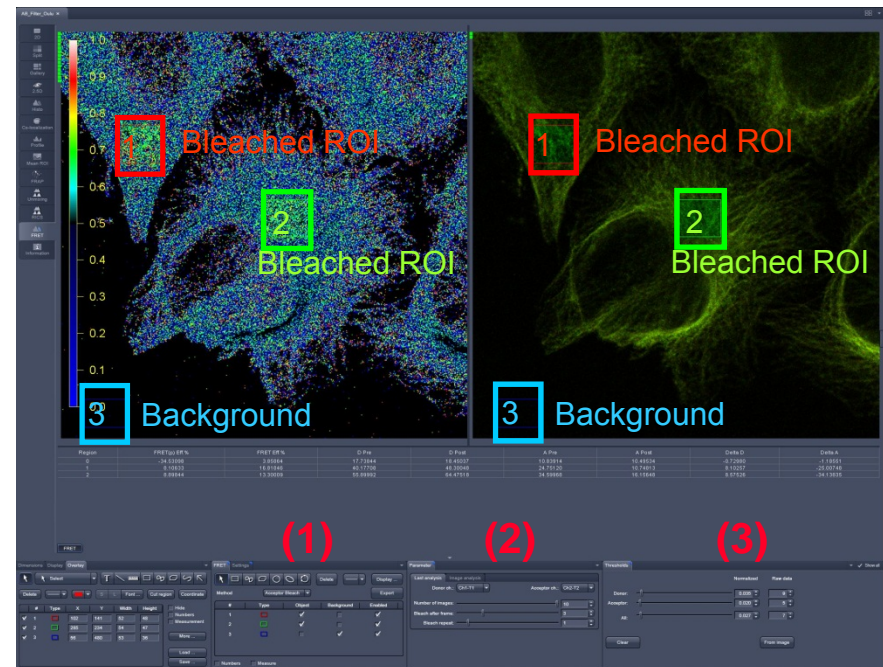
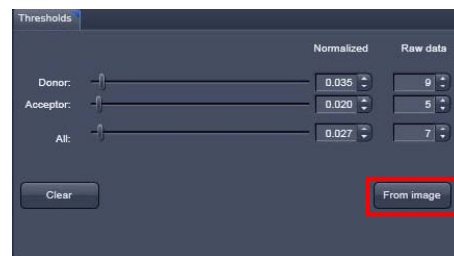
(2) Parameter

- Define acquisition and bleaching parameter



(3) Thresholds

- From image



(4) Data export

Region	FRET(p) Eff %	FRET Eff %	D Pre	D Post
0	-34.53886	3.85864	17.73844	18.45037
1	8.10633	16.81848	40.17708	48.30048
2	8.89844	13.3009	55.89992	64.47518

A Pre	A Post	Delta D	Delta A
10.83914	10.49534	-0.72980	-1.18551
24.75120	10.74813	8.10257	-25.00748
34.59968	16.15648	8.57526	-34.13835

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FRET Detection: Sensitized Emission

Principle of Sensitized Emission

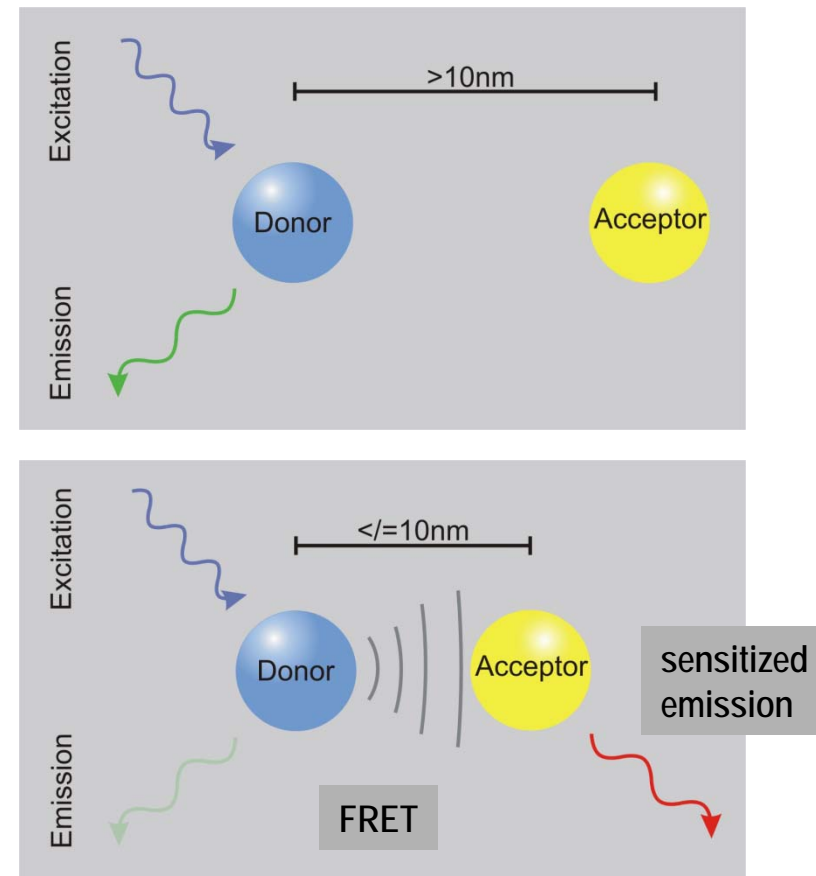


Employing FRET (Fluorescence resonance energy transfer) as a molecular ruler in microscopy

FRET is a non-radiative transfer of an excited state from one fluorophore (donor) to another (acceptor).

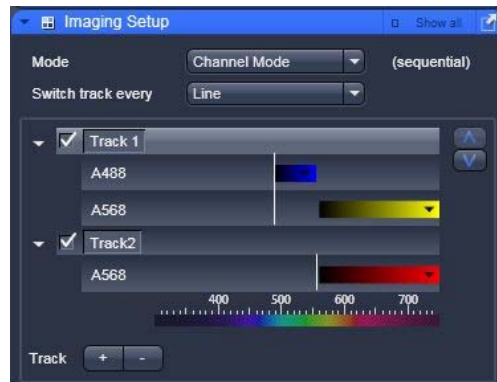
FRET occurs if donor and acceptor are in close proximity (1-10 nm).

FRET permits microscopic proximity assays at the molecular level!

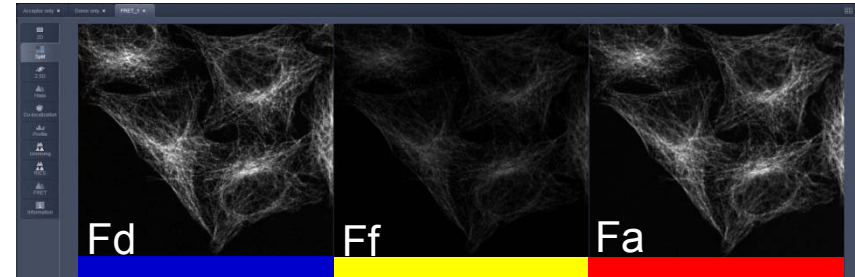


FRET Detection: Sensitized Emission

ZEN: Data evaluation in ZEN 2010 using FRET



FRET



(1) Imaging Setup

- Multitrack Channel Mode
- Linewise switch (fast)
 - (1) Track1: Donor (blue) + FRET (yellow)
 - (2) Track2: Acceptor (red)

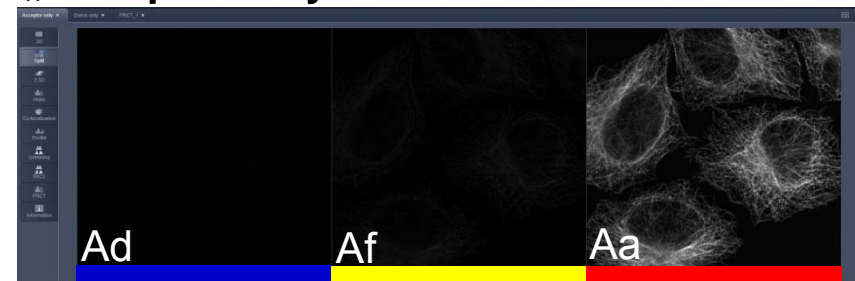
(2) Acquire images of FRET sample and references with the same settings

- Avoid noisy images (→Ratios)
- Adjust settings for FRET sample
- Acquire references with the same acquisition parameters

„Donor only“



„Acceptor only“

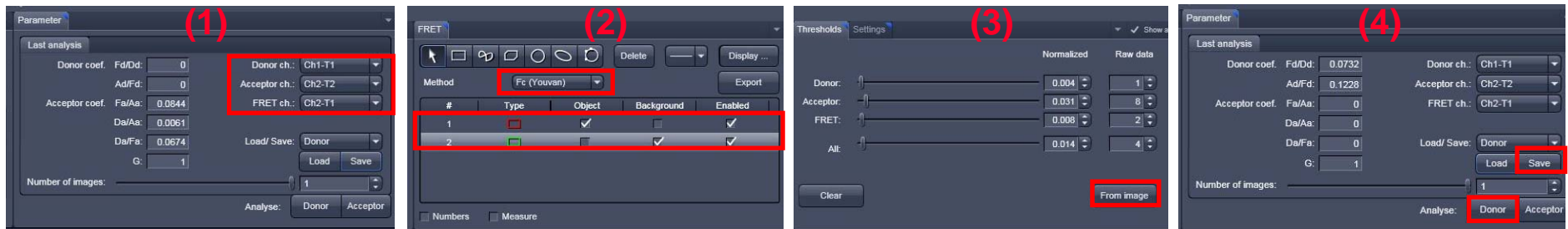


FRET Detection: Sensitized Emission

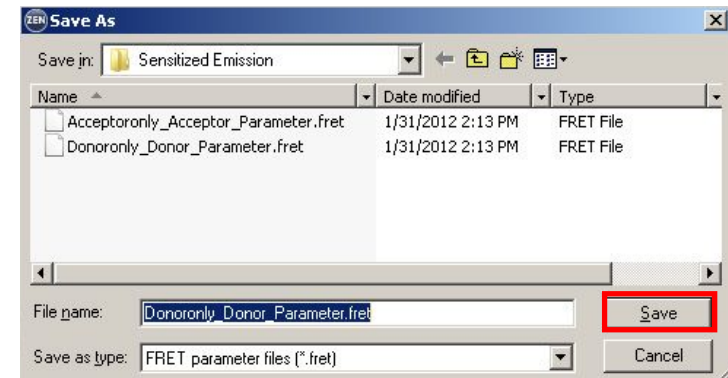
ZEN: Data evaluation in ZEN 2010 using FRET



(A) Workflow „Donor only“



- (1) **Parameter** Assign channels (→ Donor, → Acceptor, → FRET)
- (2) **FRET** Method Fc Youvan
Define ROIs / Enable Object(s) and Background
- (3) **Thresholds** „From image“
- (4) **Parameter** Analyse „Donor“
Save „Donor_Parameter.fret“

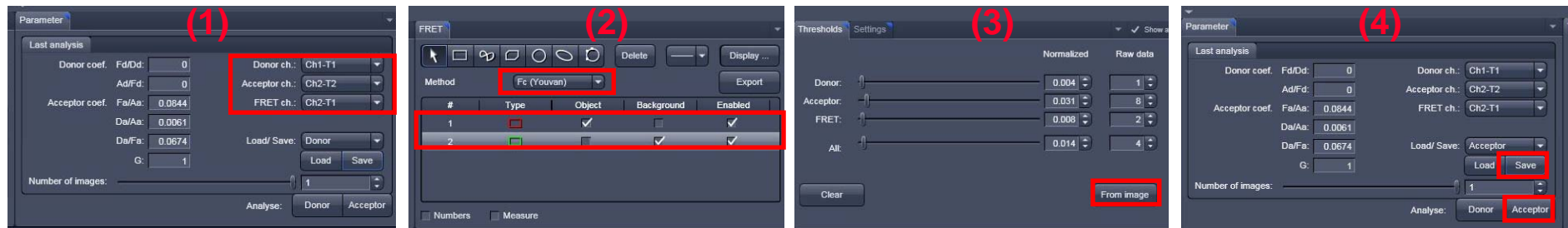


FRET Detection: Sensitized Emission

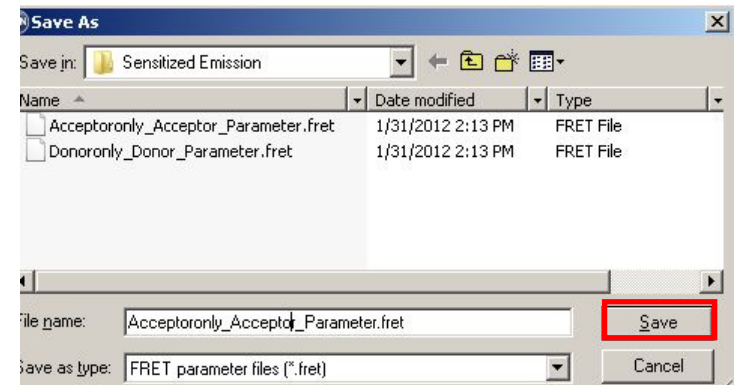
ZEN: Data evaluation in ZEN 2010 using FRET



(B) Workflow „Acceptor only“



- (1) **Parameter** Assign channels (→ Donor, → Acceptor, → FRET)
- (2) **FRET** Method Fc Youvan
Define ROIs / Enable Object(s) and Background
- (3) **Thresholds** „From image“
- (4) **Parameter** Analyse „Acceptor“
Save „Acceptor_Parameter.fret“

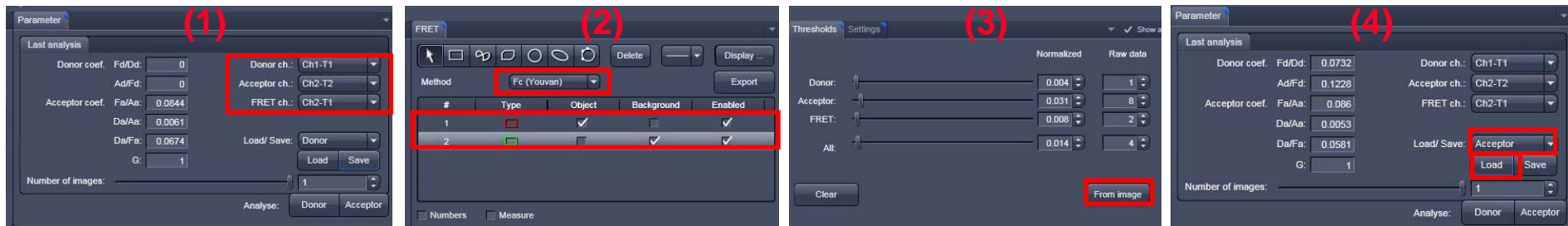


FRET Detection: Sensitized Emission

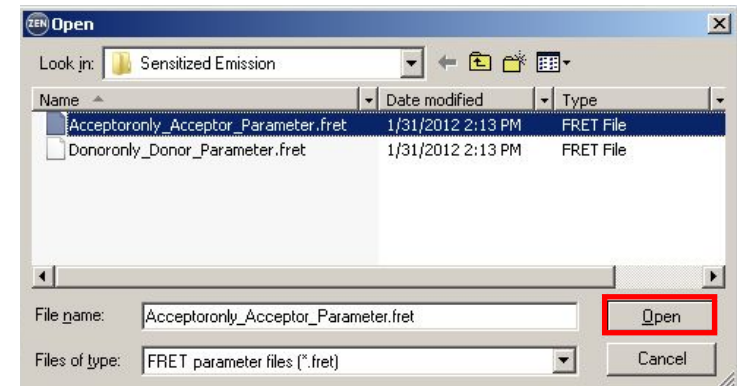
ZEN: Data evaluation in ZEN 2010 using FRET



(C) Workflow „FRET“



- (1) **Parameter** Assign channels (→ Donor, → Acceptor, → FRET)
- (2) **FRET** Method Fc Youvan
Define ROIs / Enable Object(s) and Background
- (3) **Thresholds** „From image“
- (4) **Parameter** Load „Donor“
Load „Acceptor“ (shown)



FRET Detection: Sensitized Emission

Quantifying FRET: Sensitized Emission



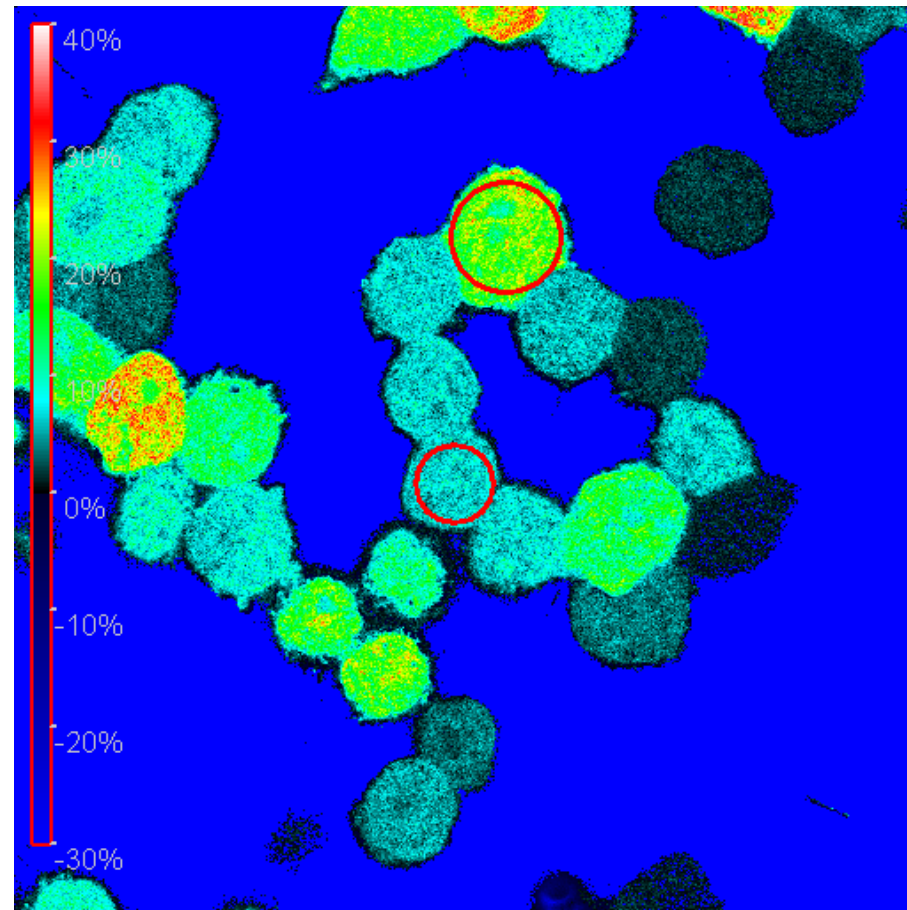
Method 1

Fc (FRETcorrected)

D.C. Youvan et al. 1997

This method calculates the absolute intensity of sensitized emission in the FRET channel as F_c . Therefore, high F_c values occur at high molecular concentrations.

$$F_c = F_f - [Donor\ corr.] - [Acc.\ corr.]$$



FRET Microscopy with LSM Systems

Quantifying FRET: Sensitized Emission



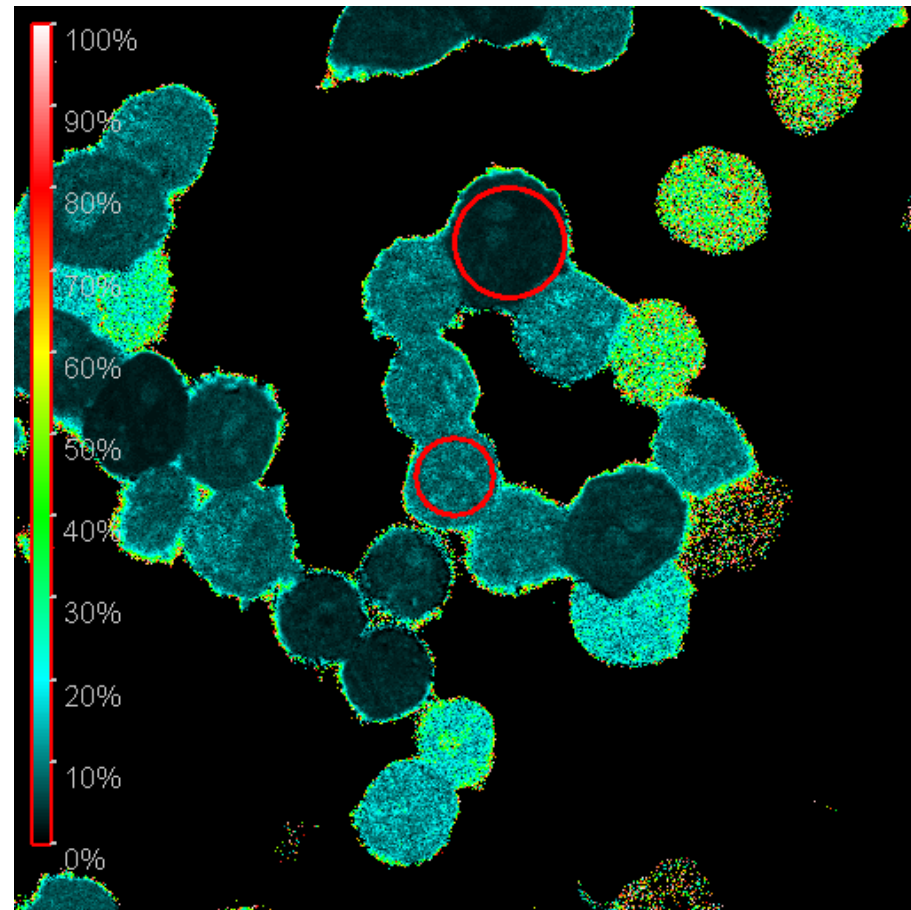
Method 2

Fn (FRET net)

G.W. Gordon et al. 1998

This method emphasizes FRET occurring at low concentrations of FRET pairs.

$$F_n = \frac{F_f - [Donor\ corr.] - [Acc.\ corr.]}{G \cdot F_d \cdot F_a}$$



FRET Microscopy with LSM Systems

Quantifying FRET: Sensitized Emission



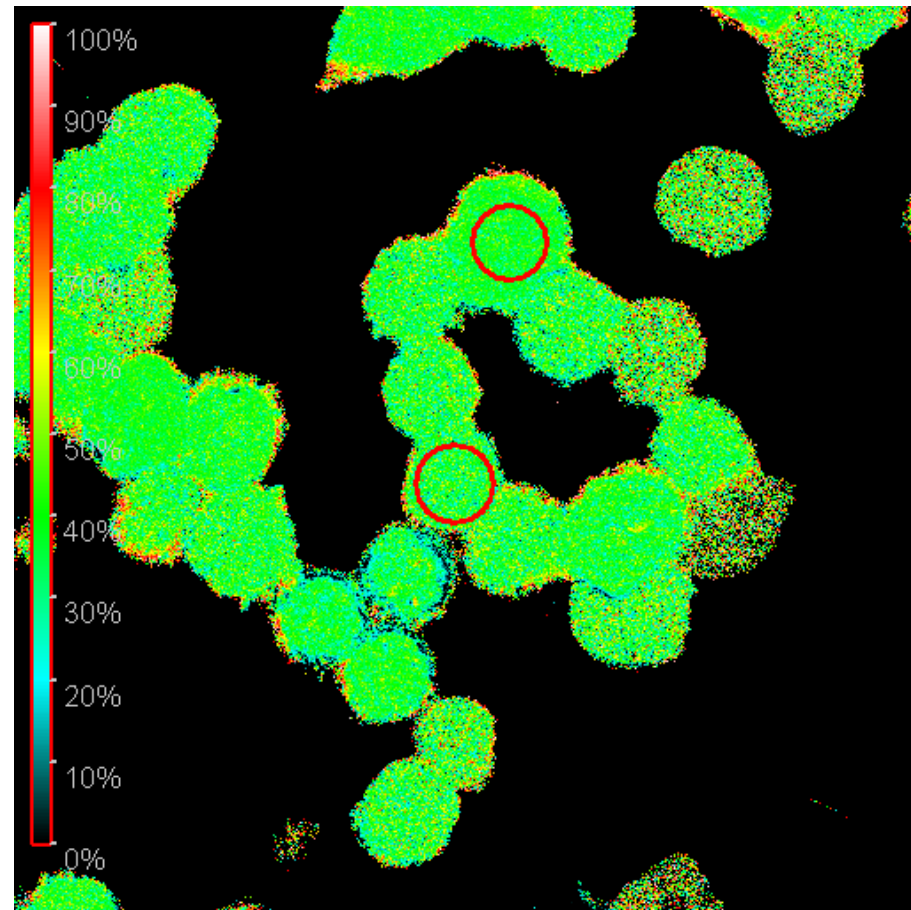
Method 3

NF (normalized FRET)

X. Xia et al. 2001

NF provides FRET values normalized for concentrations of FRET pairs.

$$NF = \frac{Ff - [Donor\ corr.] - [Acc.\ corr.]}{\sqrt{G \cdot Fd \cdot Fa}}$$



Sensitized Emission – The 3 (main) Methods

Method 1: F_c (FRET corrected) D.C. **Youvan** et al. 1997
 Dependend on Donor / Acceptor concentration. If its less → no FRET Signal

$$F_c = F_f - [Donor \ corr.] - [Acc. \ corr.]$$

Method 2: F_n (FRET net) G.W. **Gordon** et al. 1998
 Facilitates getting FRET Signals also if Donor / Acceptor signals are weak

$$F_n = \frac{F_f - [Donor \ corr.] - [Acc. \ corr.]}{G \cdot F_d \cdot F_a}$$

Method 3: NF (normalized FRET) X. **Xia** et al. 2001
 Independent of Donor und Akzeptor concentration

$$NF = \frac{F_f - [Donor \ corr.] - [Acc. \ corr.]}{\sqrt{G \cdot F_d \cdot F_a}}$$